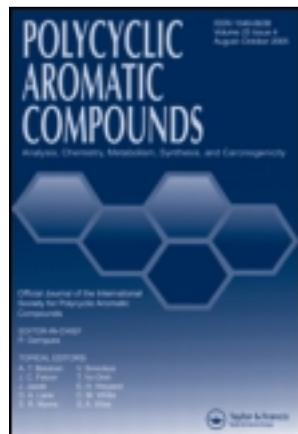


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The Effect of Local Cooking Methods on Polycyclic Aromatic Hydrocarbons (PAHs) Contents in Beef, Goat Meat, and Pork as Potential Sources of Human Exposure in Kisumu City, Kenya

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Roasted meat is known to be a major source of human exposure to PAHs. The contribution of direct-heat charcoal-roasted, electric-oven grilled, and shallow-pan fried meat to human exposure in Kisumu City was not known although the three modes of cooking meat are very prevalent. This study analyzed the concentrations of the PAHs in raw beef, goat meat, and pork, investigated the effect of direct-heat charcoal roasting, electric-oven grilling, and shallow-pan frying on these concentrations, and compared their concentration levels with international standards for foods in order to assess the potential risks to consumers. Samples were taken from three popular meat-roasting hotels within Kisumu City, Kenya. Extraction of PAHs was done using liquid-liquid partition after saponification with alcoholic potassium hydroxide followed by clean-up on a silica gel column and final analysis by gas chromatography-mass spectrometry (GC-MS). Roasting and shallow-pan frying introduced new PAHs and significantly

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($P \leq 0.05$) increased the concentrations of those existing in raw meat. Direct-heat charcoal roast beef had 5 new PAHs and a total mean PAH content of $17.88 \mu\text{g}/\text{kg}$, compared with a mean of $1.39 \mu\text{g}/\text{kg}$ for raw beef, with the potent dibenz(a,h)anthracene also being detected. Direct-heat charcoal roasted goat meat had three new PAHs and a total mean PAH content of $4.77 \mu\text{g}/\text{kg}$, compared with a mean of $2.13 \mu\text{g}/\text{kg}$ in raw meat, with the potent benzo(a)pyrene concentration being 8.84% of the total mean PAH. Fried pork had 7 new PAHs and a total mean PAH content of $3.47 \mu\text{g}/\text{kg}$, compared with a mean total of $0.17 \mu\text{g}/\text{kg}$, detected in the raw meat. Roast beef had the highest individual PAH concentration ($5.03 \mu\text{g}/\text{kg}$) and highest total PAHs concentration ($17.88 \mu\text{g}/\text{kg}$), both being higher than acceptable EU limits. The PAHs from local raw and cooked meat were characterized and quantified for the first time in Kisumu City and the study therefore provided the needed baseline data on PAHs in raw and cooked meat.

Key Words: frying, Kenya, Kisumu city, meat, polycyclic aromatic hydrocarbons, roasting

INTRODUCTION

PAHs are of great concern in environmental monitoring because they are known or suspected carcinogens and/or toxicants. The United States Environmental Protection Agency (USEPA) lists 16 PAHs as priority pollutants that need to be periodically monitored in the environment because of their known carcinogenicity. The USEPA (1) developed an oral “virtually safe dose” (VSD) for benzo(a)pyrene of $0.14 \text{ ng}/\text{kg}$ body weight/day to monitor the carcinogenic risk it poses. Cancer incidence in Kenya is increasing (data from Nairobi Cancer Registry and verbal reports from practicing physicians; Kakamega Hospice Registry) as reported by Lisouza et al. (2) and now numbers among the top 10 causes of mortality (2, 3). However, the cause of the increase in cancer incidence and prevalence is not known. The population may be exposing themselves to carcinogens through food by eating contaminated fish (4–6) and roasted, grilled, and fried meat (7–12). Human exposure may also be through smoke (smoking tobacco, smoke from burning of wood, and petroleum fuel (13)) and emissions from creosote treated wood products (14), foods from soils contaminated with PAHs, and industrial workplace exposure (15).

Other potential sources of PAHs in the environment exist in Kisumu City with the major point sources including petroleum fuel spillages, Kenya Pipeline Company depot runoff, car wash activities at the shore, oil spills from vessels at the Pier and Yatch Club, mechanical workshops (*Jua Kali* sheds), and petrol-station runoff (16). Mobile sources include motor vehicle exhaust (17) and consumer products waste dumped into the lake. Diffuse sources include asphalt roads and road tar, fires of all types (municipal garbage incineration and burning of sugar cane from the surrounding sugar belt) and biomass energy combustion (2, 4, 18), agricultural runoff, and natural alteration of organic matter (19). It is not known how much these sources contribute to the PAH exposure as their levels are neither known nor periodically monitored in

meat products which are main components of diet in the area. Recent studies have indicated that relatively high levels of PAHs are present in sediment and water in Winam Gulf of Lake Victoria where the city gets its water from (16).

The aim of this study was to analyze the concentration levels of PAHs in raw and cooked meat from selected hotels in Kisumu City in order to provide baseline data and knowledge on potential exposure in humans. Various cooking methods are used in Kisumu City for meat preparation. The choice of a particular method depends on the type of establishment, income levels of the inhabitants, the available cooking equipment or apparatus, and the type of fuel used. The cooking methods targeted in this study were open-air direct-heat charcoal oven roasting and electric-oven grilling (for beef and goat meat) and shallow-pan frying (for pork). These are the local meat cooking methods used in the hotels that were selected for the study. Pork is not served roasted and, as such, roasting pork would not provide a true potential source for PAH exposure in humans in our case. The pork roasting method was therefore not included in the sampling design in the study.

MATERIALS AND METHODS

General Purpose Grade (GPR) methanol and diethyl ether from Kobian (K) Ltd, Nairobi, were distilled in the laboratory prior to use for extraction. Analytical HPLC grade n-hexane, 95% pure, was obtained from Kobian (K) Ltd, Nairobi. Potassium hydroxide, sodium sulphide ($\text{Na}_2\text{S}\cdot 9\text{H}_2\text{O}$), anhydrous sodium sulphate (Na_2SO_4), and silica gel (230–400 mesh) were pure analytical grade and were obtained from Kobian (K) Ltd, Nairobi. Pure nitrogen and helium for gas chromatography-mass spectrometric (GC-MS) analysis were obtained from East African Oxygen Company, Kisumu, Kenya.

Sampling

Samples of raw, open-air direct-heat charcoal- and electric-oven grill roasted and shallow-pan fried meat were taken in triplicates from various meat-roasting places in Kisumu City ($0^\circ 6'S$, $34^\circ 45'E$) located on the North tip of Winam Gulf, Lake Victoria, Kenya (Figure 1). Kisumu City has a population of 504,000 and the most common sources of protein for the local population include fish, beef, mutton, goat meat, and pork. The specific sampling sites were Highway Inn (on Kisumu-Kakamega Highway) for goat meat and beef, Apok Inn (on Kisumu-Nairobi Highway) for beef, and Kisumu Hotel (in the city center) for pork. A sample of raw meat (1 kg) was taken and divided into halves. The first half was packed for laboratory analysis while raw and the second half was open-air charcoal roasted using direct heat (Highway Inn), electric-oven grilled (Apok Inn), or shallow-pan fried (Kisumu Hotel) as necessary before

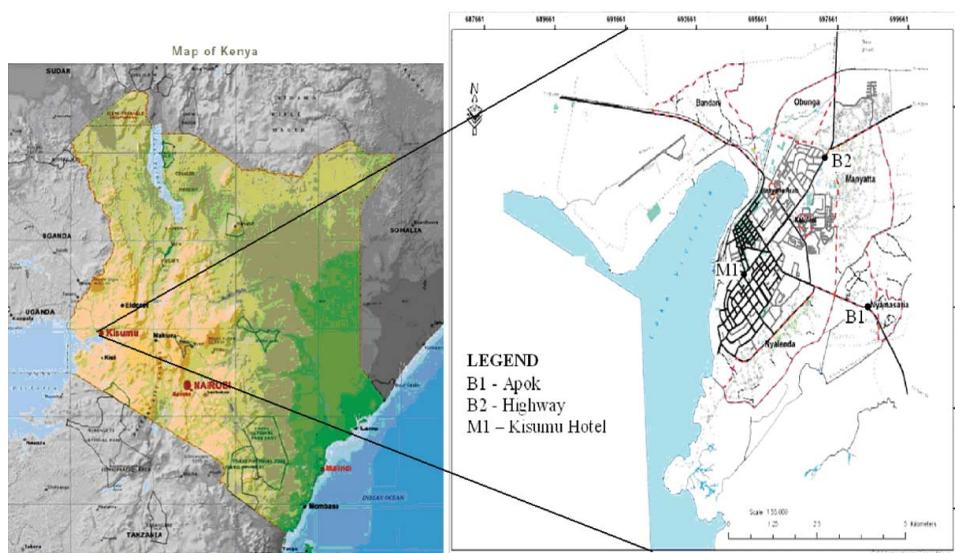


Figure 1: Map of Kenya showing Kisumu City and Bay ($0^{\circ}6'S$, $34^{\circ}45'E$) with sampling locations (color figure available online).

packing. The samples were wrapped in pieces of pre-cleaned aluminum foil and placed in an ice-cold container for transport to the laboratory.

Extraction and GC-MS Analysis of PAHs

Extraction of PAHs from raw meat followed a modified method described by Takatsuki et al. (4) where samples of meat were cut into small pieces and homogenized in a blender; 50 g of the homogenized sample was saponified using a mixture of methanol (200 mL), 50% aqueous potassium hydroxide (35 mL) and sodium sulphide (2 g) and then refluxed on a water bath for 2 h. The mixture was cooled to 40°C and 150 mL of n-hexane added in small portions, with occasional swirling. The mixture was poured into 150 mL de-ionized water in a 500-mL glass separation funnel, the flask rinsed twice with 10 mL methanol, and the rinses added to the mixture. The separation funnel was gently and adequately shaken and then set to separate. The aqueous layer was extracted twice with 150 mL and 100 mL portions of n-hexane, respectively. All the n-hexane extracts were combined, washed with 100 mL de-ionized water, dried over anhydrous sodium sulphate, transferred to a rotary evaporator, and then concentrated to about 3–5 mL.

For clean-up and separation, a chromatographic glass column (1 cm diameter) was packed with 8 g silica gel (in a slurry) and 3 g anhydrous Na_2SO_4 , added on top. The column was covered with aluminum foil to protect it from light and washed with 30 mL n-hexane; the solvent was then drained to just above the slurry. The concentrated extract was transferred to the column.

Clean up and elution of PAHs was done slowly under gravity with 150 mL volume of 10% diethyl ether in n-hexane. The extract was evaporated to 1–2 mL, in a rotary evaporator, then under a mild stream of nitrogen to dryness with gentle warming and the residue was taken up in 1 mL pentane (Sigma, St. Louis, MO) and stored in sealed vials for analysis.

The samples were analyzed by GC-MS, a 7890A stand-alone GC (Agilent Technologies, Inc., Beijing, China) and a 5975 C MSD (Agilent Technologies, Inc., Santa Clara, CA). The conditions were: inlet temp 270°C, transfer line temp of 280°C, and column oven temperature programmed from 35–285°C initially for 5 min then 10°C/min to 280°C for 10.5 min and then at 5°C/min to 285°C for 29.9 min. The GC had HP-5 MS low bleed capillary column (30 m × 0.25 mm i.d., 0.25- μ m) (Restek, Bellefonte, PA). Helium was the carrier gas (flow rate 1.25 mL/min). The MSD ion source temperature was 250°C and quadrupole temperature of 180°C. Electron impact mass spectra were obtained at an acceleration energy of 70 eV. Fragment ions were analyzed over 40–550 m/z mass range in the full scan mode. A 1.0 μ L aliquot of extract was automatically injected in the split/splitless mode using an auto sampler 7683 (Agilent Technologies, Inc., Beijing, China). The filament delay time was set as 3.3 min. Library –MS searches were done using NIST/EPA/NIH Mass Spectral Library (NIST 05) and NIST Mass Spectral Search Program Version 2.0d.

External PAH standards (PAH Mix 9; Dr. Ehrenstorfer mbH, Augsburg, Germany) were used in different dilutions to come up with calibration curves which were used to relate the various concentrations to peak areas. Calibration standards of 5 concentration levels—0.1 mg/L, 0.5 mg/L, 1 mg/L, 5 mg/L, and 10 mg/L—were prepared in HPLC-grade pentane. This was used to give the peak retention times for identification and the calibration curves for quantification. To evaluate the recovery of PAH and to account for matrix effects in the GC–MS chromatograms, spiked control samples were analyzed through the same procedure. The results obtained were then corrected based on percentage recovery for each PAH (Table 1).

Statistical Analysis

The results were analyzed using a factorial two design with PAH type as the main treatment and meat type (raw and cooked) as a sub-treatment. The results obtained were then processed for ANOVA using INSTAT software. For beef the sub-treatments were meat type and location.

RESULTS AND DISCUSSION

It should be noted that these results are based on a method that is not very recent (4). However, there are more recent and more sensitive methods for

Table 1: PAHs recovery in meat samples

| PAH type | % recovery \pm standard deviation |
|------------------------|-------------------------------------|
| Naphthalene | 83.5 \pm 11.5 |
| Acenaphthylene | 85.0 \pm 7.9 |
| Acenaphthene | 80.0 \pm 12.3 |
| Fluorene | 86.3 \pm 6.9 |
| Phenanthrene | 92.5 \pm 1.5 |
| Anthracene | 96.5 \pm 1.4 |
| Fluoranthene | 91.6 \pm 11.4 |
| Pyrene | 92.4 \pm 2.5 |
| Chrysene | 76.2 \pm 12.1 |
| Benz(a)anthracene | 72.7 \pm 18.0 |
| Benzo(b)fluoranthene | 98.3 \pm 2.1 |
| Benzo(k)fluoranthene | 87.0 \pm 19.1 |
| Dibenz(a,h)anthracene | 87.9 \pm 12.7 |
| Benzo(a)pyrene | 94.4 \pm 5.6 |
| Indeno(1,2,3-cd)pyrene | 91.7 \pm 2.9 |
| Benzo(g,h,i)perylene | 76.8 \pm 2.5 |

determination of PAHs, for example, Gosetti et al. method (21). However, the use of GC-MS and the quality control checks done during the analysis support the accuracy of our method.

PAHs in Raw Meat

Raw beef was found to contain 7 of the 16 priority PAHs, as shown in Table 2. The sample from The Highway Inn contained 7 of the 16 priority PAHs while beef from The Apok Inn exhibited three of the priority PAHs. Average background values are usually in the range of 0.01–1 $\mu\text{g}/\text{kg}$ in uncooked foods (20, 24) and the values obtained at Highway are above that range.

Apok and Highway obtain their beef from the same slaughterhouse. At Highway, the beef is kept in the open in close proximity to the roasting place prior to cooking. This could expose the raw beef to ambient PAHs originating from the roasting place (2) which could get adsorbed onto the surface of the meat. However, at Apok, the meat is kept at a butchery which is located in a different room from the kitchen where roasting is done.

Raw goat meat was found to contain 9 of the 16 priority PAHs (Table 2). The PAHs were within the average background values for uncooked foods (20) with the exception of anthracene which had the highest mean PAH concentration recorded. However, the total mean PAH concentration was well above the average background value in uncooked foods (20). Raw pork contained 2 of the 16 priority PAHs (Table 2). The values are within the average background values for uncooked foods (20).

Table 2: The mean PAH concentration in raw meat (\pm standard deviation in $\mu\text{g}/\text{kg ww}$)

| PAH | BEEF | | | GOAT MEAT | | PORK | | MEAN PAH |
|-----------------------|-------------------|-------------------|-------------------|-------------------|-------------------|-------------------|-------------------|--------------|
| | Apok | Highway | | Mean | Highway | | Kisumu Hotel | All sites |
| | | Highway | Mean | | Highway | Mean | | |
| Naphthalene | 0.087 \pm 0.003 | 0.118 \pm 0.015 | 0.102 \pm 0.019 | 0.105 \pm 0.007 | nd | nd | nd | 0.077 |
| Acenaphthylene | 0.070 \pm 0.001 | 0.124 \pm 0.004 | 0.097 \pm 0.030 | 0.031 \pm 0.004 | 0.097 \pm 0.005 | 0.097 \pm 0.005 | 0.097 \pm 0.005 | 0.081 |
| Acenaphthene | 0.053 \pm 0.001 | 0.086 \pm 0.002 | 0.069 \pm 0.018 | 0.074 \pm 0.004 | 0.072 \pm 0.003 | 0.072 \pm 0.003 | 0.072 \pm 0.003 | 0.071 |
| Fluorene | nd | 0.310 \pm 0.009 | 0.155 \pm 0.170 | 0.229 \pm 0.020 | nd | nd | nd | 0.337 |
| Anthracene | nd | 0.870 \pm 0.001 | 0.435 \pm 0.477 | 1.033 \pm 0.045 | nd | nd | nd | 0.476 |
| Pyrene | nd | 0.934 \pm 0.010 | 0.467 \pm 0.512 | nd | nd | nd | nd | 0.234 |
| Benz(k)fluoranthene | nd | nd | nd | 0.332 \pm 0.009 | nd | nd | nd | 0.083 |
| Indeno(1,2,3-cd)P | nd | 0.097 \pm 0.002 | 0.067 \pm 0.073 | 0.097 \pm 0.003 | nd | nd | nd | 0.058 |
| Db(a,h)A | nd | nd | nd | 0.102 \pm 0.022 | nd | nd | nd | 0.025 |
| Benzo(G,h,i)P | nd | nd | nd | 0.127 \pm 0.011 | nd | nd | nd | 0.032 |
| Total mean PAH | 0.210 | 2.539 | 1.392 | 2.130 | 0.169 | 0.169 | 0.169 | 1.474 |

nd = not detected/below detect limit of the machine (detection limit = 10exp-12g) and assumed to be zero in statistics; Indeno(1,2,3-cd)P = Indeno(1,2,3-cd)pyrene; Db(a,h)A = Dibenz(a,h)anthracene; Benzo(G,h,i)P = Benzo(G,h,i)perylene

Effect of Direct-Heat Charcoal Roasting on PAH Concentrations in Beef

Roasting increased the concentrations of all the PAHs and also introduced new PAHs (Table 3). At Apok, the new PAHs introduced included fluorene, phenanthrene, anthracene, pyrene, benzo(k)fluoranthene, and indeno(1,2,3-cd)pyrene while the new ones at Highway included dibenz(a,h)anthracene, benzo(k)fluoranthene, and benzo(ghi)perylene. Therefore, it is a dynamic process where some PAHs are burnt out during roasting while new ones are generated (Table 3). This is comparable to what is found in literature. Janoszka *et al.* (10) who studied roast meats in Poland found that the total PAH content was within the range 2.43–16.10 ng/g meat while Reinik *et al.* (12) found 16 µg/kg in smoked meat and ham as part of Estonian Food Safety Monitoring. Martonell *et al.* (11), however, reported higher levels at 33.99 µg/kg in meat and meat products in a population exposure study of Catalonia, Spain. In this study, the concentrations of individual PAHs were also significantly different with respect to location with Highway showing higher levels than Apok ($P \leq 0.05$). This could be due to the fact that Apok used an indoor electric-oven grill to prepare their meat, which would yield less PAHs, while Highway used open-air charcoal stove which could perpetuate production of more PAHs (2, 22).

Table 3: Effect of roasting on the PAH concentration in beef (in µg/kg ww)

| PAH | Apok | Highway | Roast Mean | Raw Mean | LSD (P = 0.05) | CV(%) |
|-----------------------|-------------|--------------|---------------|--------------|----------------|--------|
| Naphthalene | 0.113 | 0.233 | 0.173 | 0.102 | 0.016 | 19.70 |
| Acenaphthylene | 0.087 | 0.256 | 0.172 | 0.097 | 0.019 | 24.80 |
| Acenaphthene | 0.065 | 0.193 | 0.129 | 0.069 | 0.016 | 27.40 |
| Fluorene | 0.255 | 1.604 | 0.930 | 0.155 | 0.173 | 55.30 |
| Phenanthrene | 0.223 | nd | 0.112 | nd | 0.037 | 115.80 |
| Anthracene | 0.832 | 4.804 | 2.818 | 0.435 | 0.540 | 57.80 |
| Fluoranthene | nd | 1.270 | 0.635 | nd | 0.212 | 115.60 |
| Pyrene | 0.395 | 3.648 | 2.021 | 0.467 | 0.387 | 53.80 |
| B(k)F | 0.143 | 0.202 | 0.172 | nd | 0.010 | 20.10 |
| Indeno(1,2,3-cd)P | 0.097 | 0.253 | 0.175 | 0.067 | 0.006 | 9.10 |
| Db(a,h)A | nd | 5.032 | 2.516 | nd | 0.839 | 115.50 |
| Benz(g,h,i)P | nd | 0.385 | 0.193 | nd | 0.064 | 115.50 |
| Total mean PAH | 2.21 | 17.88 | 10.046 | 1.392 | | |

nd = not detected/below detect limit of the machine (detection limit = 10×10^{-12} g) and assumed to be zero in statistics; CV: Coefficient of variation; LSD: Least square deviation; B(k)F = Benzo(k)fluoranthene; Indeno(1,2,3-cd)P = Indeno(1,2,3-cd)pyrene; Db(a,h)A = Dibenz(a,h)anthracene; Benzo(g,h,i)P = Benzo(g,h,i)perylene; Note: Apok: electric-oven grill used, Highway: charcoal stove used.

Effect of Direct-Heat Charcoal Roasting on PAH Concentrations in Goat Meat

Acenaphthene, anthracene, indeno(1,2,3-cd)pyrene, and dibenz(a,h)anthracene concentrations were found to be significantly higher ($P \leq 0.05$) in roasted than in raw goat meat (Table 4). Twelve of the 16 priority PAHs were found in roasted goat meat as compared to the nine found in raw goat meat. The new PAHs generated included phenanthrene, fluoranthene and the potent benzo(a)pyrene whose percentage in comparison to total PAH concentration was 8.84% (Table 4). This is relatively low as compared to the range 2.43–16.10 ng/g found by Janoszka et al. (10) in roast meats in Poland; 16 $\mu\text{g}/\text{kg}$ in smoked meat and ham found by Reinik et al. (12), as part of Estonian Food Safety Monitoring, and 33.99 $\mu\text{g}/\text{kg}$ in meat and meat products found by Martonell et al. (11) in a population exposure study of Catalonia, Spain. This shows that the local roasting methods are safer than these smoking methods reported in literature. However, the type of meat roasted influences the PAHs concentrations as higher total PAH concentrations were recorded in direct-heat charcoal roast beef than in direct-heat charcoal roasted goat meat.

Effect of Shallow-Pan Frying on PAH Concentrations in Pork

Frying increased the number of PAHs detected in raw pork from two to nine. However, the concentration of the PAHs existing in raw pork were not significantly ($P \leq 0.05$) increased by frying (Table 5). The new

Table 4: Effect of charcoal roasting on the PAH concentrations in goat meat ($\mu\text{g}/\text{kg}$ ww)

| PAH | Roast Mean | Raw Mean | LSD ($P = 0.05$) | CV (%) |
|------------------------|--------------|--------------|--------------------|--------|
| Naphthalene | 0.308 | 0.105 | 0.026 | 3.55 |
| Acenaphthylene | 0.126 | 0.031 | 0.007 | 2.56 |
| Acenaphthene | 0.092 | 0.074 | 0.022 | 7.53 |
| Fluorene | 0.892 | 0.229 | 0.073 | 3.50 |
| Phenanthrene | 0.207 | nd | 0.020 | 5.54 |
| anthracene | 1.670 | 1.033 | 1.305 | 31.35 |
| Fluoranthene | 0.251 | nd | 0.035 | 7.86 |
| Benz(k)fluoranthene | 0.410 | 0.332 | 0.024 | 1.86 |
| Benzo(a)pyrene | 0.422 | nd | 0.011 | 1.44 |
| Indeno(1,2,3-cd)pyrene | 0.121 | 0.097 | 0.100 | 2.54 |
| Dibenz(a,h)anthracene | 0.113 | 0.102 | 0.069 | 18.24 |
| Benz(g,h,i)perylene | 0.164 | 0.127 | 0.020 | 3.99 |
| Total mean PAH | 4.774 | 2.130 | | |

nd = not detected/below detect limit of the machine (detection limit = $10 \times 10^{-12}\text{g}$) and assumed to be zero in statistics

Table 5: Effect of shallow-pan frying on PAH concentrations in pork ($\mu\text{g}/\text{kg}$)

| PAH | Fried Mean | Raw Mean | LSD (P = 0.05) | CV (%) |
|-----------------------|--------------|--------------|----------------|--------|
| Naphthalene | 0.074 | nd | 0.011 | 8.44 |
| Acenaphthylene | 0.325 | 0.097 | 0.025 | 3.37 |
| Acenaphthene | 0.367 | 0.072 | 0.055 | 7.07 |
| Fluorene | 0.341 | nd | 0.039 | 6.51 |
| Anthracene | 0.543 | nd | 0.047 | 4.98 |
| Fluoranthene | 0.491 | nd | 0.020 | 2.26 |
| Benzo(a)pyrene | 0.701 | nd | 1.295 | 105.10 |
| Benz(k)fluoranthene | 0.237 | nd | 0.063 | 15.06 |
| Indeno(1,2,3-cd)P | 0.386 | nd | 0.069 | 10.17 |
| Total mean PAH | 3.465 | 0.169 | | |

nd = not detected/below detect limit of the machine (detection limit = $10\text{exp}-12\text{g}$) and assumed to be zero in statistics; Indeno(1,2,3-cd)P = Indeno(1,2,3-cd)pyrene

PAHs introduced included fluorene, anthracene, fluoranthene, benzo(a)pyrene, benzo(k)fluoranthene and indeno(1,2,3-cd)pyrene. The concentration levels found in this study were much lower than what Janoszka et al.,(10) found showing that shallow-pan frying of pork generated less PAHs.

Comparison of the Concentration Levels of PAHs with the Standards Allowed Internationally in These Foods

The values of individual PAHs in raw beef, goat meat, and pork are lower than the maximum limits set by Commission Regulation (23) of $5 \mu\text{g}/\text{kg}$ for benzo[a]pyrene (Table 6). For the roast/fried foods studied, Highway beef has a greater potential source of exposure (total 16 USEPA PAHs) to humans followed by goat meat and pork and, the least potential source of exposure to humans after cooking is Apok beef. Highway used open-air, direct-heat, charcoal stove roasting and Apok used electric-oven grilling method. Benzo[a]pyrene was reported only in goat meat at $0.422 \mu\text{g}/\text{kg}$ which is below the maximum

Table 6: Comparison of maxima of PAH concentration range in tested foods with maximum limits set by European Commission Regulation (23) (in $\mu\text{g}/\text{kg}$)

| Sample | Beef Individual Total | | Goat Meat Individual Total | | Pork Individual Total | | ECR (2006) |
|---------------------|-----------------------|--------|----------------------------|-------|-----------------------|-------|------------|
| | PAH | PAH | PAH | PAH | PAH | PAH | |
| Raw | 0.934 | 2.105 | 1.033 | 2.201 | 0.097 | 0.169 | 2 |
| Cooked ^a | 5.032 | 17.875 | 1.670 | 4.781 | 0.701 | 3.464 | 5 |

^a roasted for beef and goat meat; fried for pork

limit set by the European Commission Regulation (23). Data on meat consumption by Kisumu population does not exist. However, data on Nairobi population's meat consumption exists (25). Gamba et al. (25) found that households in Nairobi consume a mean per adult equivalent of 15.81 kg of beef and 11.65 kg goat meat annually. Assuming that meat consumption in Kisumu follows the same pattern, then the daily consumption of goat meat is 0.032 kg per person (based on 365 days in a year) and the daily benzo(a)pyrene exposure is 13.50 ng. This implies that based on the USEPA VSD (1), one should weigh at least 96.46 kg to be safe from this level of exposure.

CONCLUSION

The PAH concentration in raw meats, both individual and total, were in the $\mu\text{g}/\text{kg}$ range. Open-air charcoal-roasting increased the PAH concentration in beef and goat meat significantly ($P \leq 0.05$) in all cases. It also introduced new PAHs in these foods. Shallow-pan frying did not increase the PAH concentration in pork significantly ($P \leq 0.05$) but introduced new PAHs. The figures for benzo(a)pyrene concentration show levels higher than USEPA virtually safe dose (VSD) to persons below 96.46 kg body weight which are indicative of significant potential human exposure to total USEPA 16 PAHs. Open-air direct-heat charcoal-roasted beef posed the highest potential for exposure followed by open-air direct-heat charcoal roasted goat meat while shallow-pan fried pork was the least potential source of exposure. There is need to minimize poor handling practices such as keeping raw meat in close proximity to roasting places/kitchens from where external and ambient sources may introduce PAHs in stored raw meat as shown in this study. Electric-oven grilling is a better method of roasting than open-air direct-heat charcoal stove roasting.

REFERENCES

1. IRIS. *Integrated Risk Information System of the United States Environmental Protection Agency*, 2002. <http://www.epa.gov/iris/> (accessed on April 30, 2012).
2. Lisouza, F. A., P. O. Owuor, and J. O. Lalah. "Variation in Indoor Levels of Polycyclic Aromatic Hydrocarbons from Burning Various Biomass Types in the Traditional Grass-Roofed Households in Western Kenya." *Environ. Pollut.* 159 (2011): 1810–5.
3. American Society of Clinical Oncology (ASCO). <http://pda.asco.org/anf/Past+Issues/January+2008/Cancer+in+Kenya?cpsexcurrchannel=1> (accessed November 21, 2010).
4. Takatsuki, K., S. Suzuki, N. Sato and I. Ushizawa. "Liquid Chromatographic Determination of Polycyclic Aromatic Hydrocarbons in Fish and Shellfish." *J. Assoc. Anal. Chem.* 68, no. 5 (1985): 324–41.
5. Eisler, R. "Polycyclic Aromatic Hydrocarbon Hazards to Fish, Wildlife, and Invertebrates: A Synoptic Review." *US Fish and Wildlife Service Biological Report* 85, no. 1.11 (1987): 81.

6. Fent, K. and R. Bätischer. "Cytochrome P₄₅₀ 1A Induction Potencies of Polycyclic Aromatic Hydrocarbons in a Fish Hepatoma Cell Line: Demonstration of Additive Interactions." *Environ. Toxicol. Chem.* 19 (2000): 2047–58.
7. Békaert, C., C. Rast, V. Ferrier, A. Bispo, M. J. Jourdain, and P. Vasseur. "Use of In Vitro (Ames and Mutatox Tests) and In Vivo (Amphibian Micronucleus Test) Assays to Assess the Genotoxicity of Leachates from a Contaminated Soil." *Org. Geochem.* 30 (1999): 953–62.
8. Bispo, A., M. J. Jourdain, and M. Jauzein. "Toxicity and Genotoxicity of Industrial Soils Polluted by Polycyclic Aromatic Hydrocarbons (PAHs)." *Org. Geochem.* 30 (1999): 947–52.
9. Madill, R. E. A., B. G. Brownlee, P. D. Josephy, and N. J. Bunce. "Comparison of the Ames Salmonella Assay for Assessing the Mutagenicity of Polycyclic Aromatic Compounds in Porewater from Athabasca Oil Sands Mature Fine Tailings." *Environ Sci Technol.* 33 (1999): 2510–6.
10. Janoszka, B., L. Warzecha, U. Błaszczuk, and D. Bodzek. "Organic Compounds Formed in Thermally Treated High-Protein Food." *Acta Chromatogr.* no. 14 (2004).
11. Martonell, I., G. Perello, R. Marti-Cid, V. Castell, J. M. Llobet, and J. L. Domingo. "PAHs in Food and Estimated PAH Intake by Population of Catalonia, Spain: Temporal Trend." *Environ. Int.* 36 (2010): 424–32.
12. Reiniik, M., T. Tamme, M. Roasto, K. Juhkam, T. Tenno, and A. Kiis. "PAHs in Meat Products and Estimated PAH Intake by Children and the General Population in Estonia." *Food Addit. Contam.* 30 (2007): 429–37.
13. Agency for Toxic Substances and Disease Registry (ATSDR). *Toxicological Profile for Polycyclic Aromatic Hydrocarbons (PAHs)* (Atlanta: U.S Department of Health and Human Services, Public Health Service, 1995).
14. Food and Environmental Hygiene Department (FEHD). *Polycyclic Aromatic Hydrocarbons in Barbecued Meat*, Risk Assessment Studies Report No. 14, Chemical Hazard Evaluation, Government of the Hong Kong Special Administrative Region, 2004.
15. Department of Environment and Heritage (DEH). *National Pollutant Inventory Substances Profile* (Queensland: Commonwealth of Australia, 2004).
16. Bowa, K. O., J. O. Lalah, and S. O. Wandiga. "Spatial and Seasonal Variations in Concentrations of Polycyclic Aromatic Hydrocarbons in Water and Sediment of Kisumu City Bay of Winam Gulf, Lake Victoria-Kenya." *Bull. Environ. Contam. Toxicol.* 83 (2009): 722–33.
17. Lalah, J. O. and P. N. Kaigwara. "Polynuclear Aromatic Compounds in Kerosene, Diesel and Unmodified Sunflower Oil and in Respective Engine Exhaust Particulate Emissions." *Toxicol. Environ. Chem.* 10, no. 1 (2005): 1–17.
18. Gu, S., A. C. Kracovec, E. R. Christensen, and R. P. vanCamp. "Source Apportionment of Polycyclic Aromatic Hydrocarbons in Dated Sediments from Black River Ohio." *Water Res.* 37 (2003): 2149–61.
19. Mitra, S. and T. S. Bianchi. "A Preliminary Assessment of Polycyclic 264 Aromatic Hydrocarbons Distribution in the Lower Mississippi 265 River and Gulf of Mexico." *Mar. Chem.* 82 (2003): 273–88.
20. Scientific Committee on Food (SCF). *Opinion of Scientific Committee on Food on the Risks to Human Health of Polycyclic Aromatic Hydrocarbons in Food*, European Commission; Health and Consumer Protection Directorate-General, Brussels, 2002.

21. Gosetti, F., U. Chiuminatto, E. Mazzucco, E. Roboti, G. Calabrese, M. C. Genaro, and E. Marengo. "Simultaneous Determination of Thirteen PAHs and Twelve Aldehydes in Cooked Food by an Automated On-Line Solid Phase Extraction and Ultra High Performance Liquid Chromatography Tandem Mass Spectrometry." *J. Chromatogr. A* 1218, no. 37 (2011): 6308–18.
22. Food and Environmental Hygiene Department (FEHD). *Polycyclic Aromatic Hydrocarbons in Barbecued Meat*. Risk Assessment Studies Report No. 14. Chemical Hazard Evaluation, The Government of the Hong Kong Special Administrative Region, 2004.
23. EU. European Commission: Commission Regulation (EC) no. 247 1881/2006, 2006.
24. National Registry of Cancer (NRC). *Polycyclic Aromatic Hydrocarbons: Evaluation of Sources and Effects* (Washington, D.C.: National Academic Press, 1983).
25. Gamba, P., D. Kariuki, and B. Gathigi. "Urban domestic consumption patterns for meat: trends and policy implications," Tegemeo Institute of Agricultural Policy Development, Egerton University, Kenya (2005).